

CLOSTRIDIAL MYONECROSIS (GAS GANGRENE)

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Rationale

For clostridial myositis and myonecrosis (gas gangrene) or spreading clostridial cellulitis with systemic toxicity (or a presumptive diagnosis of either) the preferred treatment is a combination of hyperbaric oxygen (HBO₂), surgery, and antibiotics.

Clostridial myositis and myonecrosis or gas gangrene is an acute, rapidly progressive, non-pyogenic, invasive clostridial infection of the muscles, characterized by profound toxemia, extensive edema, massive death of tissue, and a variable degree of gas production.⁽¹⁾

Gas gangrene is either an endogenous infection, caused by contamination from a clostridial focus in the body, or an exogenous infection, mostly in patients with compound and/or complicated fractures with extensive soft tissue injuries after street accidents.

The infection is caused by anaerobic, spore-forming, Gram-positive encapsulated bacilli of the genus clostridium, discovered by William H. Welch in 1891.⁽²⁾ More than 150 species of clostridium have been recognized but the most commonly isolated is *C. perfringens type A* (95%) either alone or in combination with other pathogenic clostridia, *C. novyi* (8%), *C. septicum* (4%), and *C. histolyticum*, *C. fallax*, and *C. sordelli* (1% or less of the infections).^(3,4)

A further subdivision can be made in clostridia that are toxogenic, i.e., *C. perfringens*, *C. septicum*, *C. novyi*, and clostridia that are believed to be only proteolytic, i.e., *C. histolyticum*, *C. bifermentans*, *C. sporogenes*, and *C. fallax*, which augment an infection by their proteolytic capabilities but do not cause the classical gas gangrene syndrome. *C. tertium*, *C. sphenoides*, and *C. sordelli* can be considered as contaminants. It is not known if and what these microorganisms add to the disease process. The essential role of alpha-toxin in the pathogenesis of gas gangrene was recently confirmed by Williamson and Titball,⁽⁵⁾ who developed a genetically engineered vaccine against alpha-toxin. Immunization with the C-Domain of α -toxin proved to be of value in animal experiments.⁽⁶⁾

Clostridium perfringens is not a strict anaerobe; it may grow freely in O₂ tensions of up to 30 mmHg and in a restricted manner in O₂ tensions up to 70 mmHg.⁽⁷⁾

The complete genome sequence of *C. Perfringens* has been published recently by Shimizu et al.⁽⁸⁾

The key to understanding the pathophysiology of gas gangrene is to approach it as a clinical concept, rather than a definitive bacteriologic or pathologic entity.

For the induction of gas gangrene, two conditions have to be fulfilled:

1. The presence of clostridial spores and
2. An area of lowered oxidation-reduction potential caused by circulatory failure in a local area or by extensive soft tissue damage and necrotic muscle tissue. This condition results

in an area with a low O₂ tension where clostridial spores can develop into the vegetative form.

More than 20 different clostridial exotoxins have been identified, nine of which are implicated in the local and systemic changes seen in gas gangrene; alpha-toxin, theta-toxin, kappa-toxin, mu-toxin, nu-toxin, fibrinolysin, neuraminidase, "circulating factor," and "bursting factor."⁽⁹⁻¹¹⁾

The most prevalent is the O₂-stable lecithinase-C, alpha-toxin, which is hemolytic and tissue-necrotizing. It destroys platelets and polymorphonuclear leukocytes and causes widespread capillary damage and is often lethal.⁽¹²⁾

The other toxins are ancillary to the alpha-toxin, which gives rise to hemoglobinuria, hemolysis, jaundice, anemia, tissue necrosis, renal failure, and serious systemic effects such as cardiotoxicity and brain dysfunction. The other exotoxins are synergistic and enhance the rapid spread of infection by destroying, liquefying, and dissecting healthy tissue. The clostridial organisms surround themselves with toxins. Local host defense mechanisms are abolished when the toxin production is sufficiently high. This results in fulminating tissue destruction and further clostridial growth. Alpha-toxin can be fixed to susceptible skin cells in 20-30 min, is detoxified within 2 hours after its elaboration, and causes active immunity with production of a specific antitoxin.^(10,13) The infection, however, is so progressive with continuous production of alpha-toxin that the patient dies before any immunity can develop.

Stevens et al⁽¹⁴⁾ investigated the role of theta-toxin in the pathogenesis of clostridial gas gangrene. They found evidence for the suggestion that theta-toxin in high concentrations is a potent cytolytic and promotes direct vascular injury at the site of infection. At lower concentrations, theta-toxin activates PMNs and endothelial cells, and in so doing promotes vascular injury distally by activating adherence mechanisms by PMN-dependent adherence molecules such as the integrin CD11/CD18.

The rapid tissue necrosis associated with *C. perfringens* infection is related to progressive vascular compromise orchestrated by dysregulated host cell responses induced by theta-toxin.⁽¹⁴⁾

In earlier papers, Stevens et al^(15,1) already described the lethal effects and cardiovascular effects of purified alpha- and theta-toxins from *C. perfringens*.

An extensive and updated review about the role of clostridial toxins in the pathogenesis of gas gangrene was given by Stevens and Bryant.⁽¹⁶⁾

Awad et al,⁽¹⁷⁾ showed genetic evidence for the essential role of alpha-toxin in gas gangrene.

Eaton et al⁽¹⁸⁾ have further described the crystal structure in combination with the working mechanisms of alpha toxin. In conjunction with previous findings, almost the whole working mechanism with the structure of their toxin is known now.

Stevens et al,⁽¹⁹⁾ also showed evidence that alpha- and theta-toxins differentially modulate the immune response and induce acute tissue necrosis in clostridial gas gangrene.

Much more has become known in recent years about the action and also the interaction between the various clostridial toxins in the onset and progression of gas gangrene. A very informative review on a cellular and molecular model of the pathogenesis of clostridial myonecrosis, including the above mentioned data is given by Stevens⁽¹⁾ and Titbal.⁽¹²⁾

The action of HBO₂ on clostridia (and other anaerobes) is based on the formation of O₂ free radicals in the relative absence of free radical degrading enzymes, such as superoxide dismutases, catalases, and peroxidases. Van Unnik⁽²⁰⁾ showed that an O₂ tension of 250 mmHg is necessary to stop alpha-toxin production. Although it does not kill all clostridia, it is bacteriostatic both in vivo and in vitro.⁽²⁰⁻²⁴⁾ Tissue O₂ measurements made by Schoemaker,⁽²⁵⁾ Kivisaari and Niinikoski,⁽²⁶⁾ and Sheffield⁽²⁷⁾ have shown that treatment with HBO₂ at 3.0 atm abs is required to achieve tissue partial pressures above 300 mmHg. Free-circulating toxins and/or tissue-bound toxins are not affected by high O₂ levels but they are rapidly detoxified by normal host factors.^(9,21,28,29)

If further toxin elaboration is prevented by the addition of hyperbaric oxygen, a very sick patient can rapidly be made non-toxic.

The diagnosis of clostridial myonecrosis is based primarily on clinical data, supported by the demonstration of Gram-positive rods from the fluids of the involved tissues as well as a virtual absence of leukocytes. A leukocytosis indicates a mixed infection.

Roggentin et al.⁽³⁰⁾ developed an immunoassay for rapid and specific detection of *C. perfringens*, *C. septicum*, and *C. sordelli* by determining their sialidase activity (neuraminidase) in serum and tissue homogenates. Sialidases produced by these three clostridia were bound to polyclonal antibodies raised against the respective enzymes and immobilized onto microtiter plates. Applied to nine samples from patients, there was a high correlation between the results of the immunoassay and the bacteriological analysis of the infection.⁽³⁰⁾

Scheven⁽³¹⁾ described identification of *C. perfringens* in mixed-infected clinical materials by means of a modified reversed CAMP-test.

The onset of gas gangrene may occur between 1 and 6 hours after injury or an operation and begins with severe and sudden pain in the infected area before the clinical signs appear. This seemingly disproportionate pain in a clinically still normal area must make the clinician highly suspicious for a developing gas gangrene, especially after trauma or an operation. The body temperature is initially normal but then rises very quickly. The skin overlying the wound in the early phases appears shiny and tense and then becomes dusky and progresses to a bronze discoloration. The infection can advance at a rate of 6 inches per hour. Any delay in recognition or treatment may be fatal. Hemorrhagic bullae or vesicles may also be noted. A thin, sero-sanguinolent exudate with a sickly, sweet odor is present. Swelling and edema of the infected area is pronounced. The muscles appear dark red to black or greenish. They are noncontractile, and do not bleed when cut.

The tissue gas seen on radiographs appears as feather-like figures between muscle fibers and is an early and highly characteristic sign of clostridial myonecrosis. Crepitus is usually present as well.

The acute problem in gas gangrene is not normal tissue or already necrotic tissue, but the rapidly advancing phlegmon in between, which is caused by the continuous production of alpha toxin in infected but still viable tissue. It is essential to stop alpha-toxin production as soon as possible and to continue therapy until the advance of the disease process has been clearly arrested. Since van Unnik showed that a tissue PO₂ of 250 mmHg is necessary to stop toxin production completely, the only way to achieve this is to start hyperbaric oxygen therapy as soon as possible.⁽²⁰⁾

A minimum of three to four HBO₂ treatments is necessary for this response. Treatment starts on the basis of the clinical picture and the positive Gram-stained smear of the wound fluid (without leukocytes). HBO₂ treatment stops alpha-toxin production and inhibits bacterial growth thus enabling the body to utilize its own host defense mechanisms.⁽²⁰⁻²⁴⁾

Although a three-pronged approach consisting of HBO₂, surgery, and antibiotics is essential in treating gas gangrene, initial surgery can be restricted to opening of the wound. An initial fasciotomy may be undertaken, but lengthy and extensive procedures in these very ill patients can usually be postponed, depending on how rapidly HBO₂ therapy can be initiated. Debridement of necrotic tissue can be performed between HBO₂ treatments and should be delayed until clear demarcation between dead and viable tissues can be seen.

The first clinical results in gas gangrene were remarkable, but were difficult to reproduce in the animal model.^(22,23,46)

Despite wide variations in O₂ tolerance between small and large laboratory animals and human beings, HBO₂ therapy has been used to treat experimental clostridial infections in animals. The greatest reduction in mortality in dogs was achieved by a combination of HBO₂, surgery, and antibiotics.⁽²⁴⁾ In general, studies of several investigators^(22,23,32,33,34,46) have shown that HBO₂ substantially reduced mortality and morbidity in animals following clostridial infections, when used in combination with surgery and antibiotics.

Major retrospective clinical studies indicate that the lowest morbidity and mortality are achieved with initial conservative surgery and rapid initiation of HBO₂ therapy. Results decline progressively when HBO₂ therapy is delayed. Early aggressive surgery and delayed HBO₂ treatment lead to a significantly higher mortality and morbidity than when HBO₂ is administered promptly.^(35,36,37)

Ertmann and Havemann indicate, on the basis of their experience in a series of 136 patients, treated over a twenty year period, the necessity for a combined treatment approach. However, they place surgery earlier in the protocol, sometimes after the first hyperbaric session already. All patients treated without hyperbaric oxygen or only once or twice, died.⁽³⁸⁾

The work by Brummelkamp et al.^(39,40) updated by Bakker^(41,1) totaling 409 cases of clostridial

gas gangrene showed a mortality directly related to the clostridial infection of 11.7%. All 48 patients who died did so within 26 h after the start of HBO₂ therapy. HBO₂ therapy also greatly reduced the amputation rate: only 18% required amputation post-hyperbaric therapy vs. 50-55% following primary surgery.^(4,35,36)

Hart et al.⁽⁴²⁾ reported a 17% amputation rate with combined therapeutic management. Reduced mortality rates were also demonstrated by Hart et al.,⁽⁴²⁾ Hitchcock et al.,⁽¹⁰⁾ Holland et al.,⁽⁴³⁾ Van Zijl,⁽⁴⁴⁾ and Heimbach.⁽⁴⁵⁾ Heimbach⁽¹¹⁾ showed a 5.1% mortality rate among 58 patients whose HBO₂ therapy began within the first 24 hours; these results reinforce earlier clinical trials.

Mortality in the series of Hirn⁽⁴⁶⁾ was 28%. He concluded that mortality and morbidity could be reduced if the disease is recognized early and appropriate therapy applied promptly. He recommends adequate and operative debridement, antibiotics, HBO₂, and surgical intensive care.

In experimental monomicrobial gas gangrene, the combination therapy of surgery and HBO₂ started 45 min after the inoculation of bacteria, reduced mortality to 13% compared with 38% with surgery alone. The combination therapy appeared to be especially effective in wound healing and in prevention of morbidity compared with surgical debridement alone. The effectiveness of the combination therapy was strongly time dependent.

In the multimicrobial gas gangrene model, the addition of HBO₂ to surgery tended to reduce mortality, but the difference between the groups was not statistically significant. However, the combined therapy with surgery and HBO₂ was highly effective in reducing morbidity and mortality and improving wound healing compared with surgical debridement alone.⁽⁴⁶⁾

The advantages of early HBO₂ treatment are that:

1. It is *life-saving* because less heroic surgery needs to be performed in gravely ill patients and the cessation of alpha-toxin production is rapid.
2. It is *limb and tissue-saving* because no major amputations or excisions are done prematurely (except opening of wounds). It clarifies the demarcation, so that within 24-30 hours there is a clear distinction between dead and still-living tissue. In this way, both the number and the extent of amputations are reduced.

In 1984 Peirce already concluded that the modern treatment of gas gangrene involves the simultaneous use of antibiotics, surgical debridement and hyperbaric oxygen.⁽⁴⁷⁾ He also believed, that even at that time, it would be unethical to carry out a randomized clinical study to compare these three modalities. This opinion was based on the results published until 1984.^(41,47)

Subsequent experience continues to support the approach he recommended. With the same therapy these results have been consistent over the years, and the outcome has been further improved with advanced intensive care medicine.

Utilization Review

The recommended treatment profile consists of O₂ at 3.0 atm abs pressure for 90 min, 3 times in the first 24 hours and then twice per day for the next 2-5 days. The actual decision on

termination of treatment depends on the patient's response to HBO₂ therapy. In our series⁽⁴⁾ there was no mortality after the third hyperbaric session. This is confirmed by Ertmann and Havemann.⁽³⁸⁾ If the patient remains toxic, the treatment profile needs to be extended. Utilization review is indicated after 10 treatments.

Cost Impact

Hyperbaric oxygen reduces morbidity and prevents or lowers the level of amputation necessitated by limb gas gangrene, thereby justifying the costs. HBO₂ is generally used not longer than 5-7 days.

Evidence Grading Of The Efficacy Of Treatment

The first Report of the Hyperbaric Oxygen Therapy Committee (in 1977) put gas gangrene in Category I, where HBO₂ was considered a primary therapeutic intervention to be included with surgery and antibiotics. The justification for this was: "Since 1956 the efficacy of hyperbaric oxygen in the treatment of Clostridial gas gangrene has been amply demonstrated by numerous clinical series. Both mortality and morbidity have been greatly reduced.

Reduction of morbidity, salvage of additional major joints in limb gangrene and the saving of life in severe cases justify the costs. Treatment beyond a few days is seldom if ever required."⁽⁴⁸⁾

In 1960 the randomized clinical trial was an oddity.⁽⁴⁹⁾ In gas gangrene not much has changed since then, not in pathophysiology, experimental or in the clinical results, except the way we are establishing the evidence of our results.⁽⁴⁹⁾ The microbiological background of gas gangrene is better and better understood, but that did not change our therapeutic approach. Also the results of our therapy remain remarkable constant through the years.⁽⁴⁾

The concept of Evidence Based Medicine (EBM) is dynamic rather than static. Clinical expertise and patient choices are more and more incorporated in clinical decision making⁵⁰.

We do not know of any Randomised Controlled Trial (RCT) in gas gangrene and hyperbaric oxygen therapy, because this was judged unethical already in 1984,⁽⁴⁷⁾ considering the published results and, more important, the consistency of the results through the years.

Tibbles and Edelsberg⁽⁵¹⁾ classify gas gangrene as a disease for which the weight of scientific evidence supports hyperbaric oxygen as effective adjunctive therapy. The discovery of beneficial cellular and biochemical effects strengthened the rationale for this, although they recognize the paucity of RCT's. However, this is also true for many other therapies in many indications in clinical medicine from those years (appendectomy for acute appendicitis).

Mitton and Hailey,⁽⁵²⁾ conclude from retrospective reviews and one level IV study, that there is a strong rationale for the use of hyperbaric oxygen treatment because of evidence suggesting significant reductions in both mortality and morbidity.

Heimbach⁽¹¹⁾ found more than 1200 cases of gas gangrene treated with hyperbaric oxygen in 117

articles in the literature. When we add our 600 cases (until 2003), then we can safely assume that about 2000 patients have been treated. The results in all published series⁽⁴⁾ indicate a significant reduction in mortality and morbidity by the use of adjunctive hyperbaric oxygen.

The *European Committee for Hyperbaric Medicine* has also evaluated the evidence supporting hyperbaric oxygen in the treatment of gas gangrene. The lack of RCT's place gas gangrene officially in level III (evidence of beneficial action but weakly supported), but in the light of the vast amount of experimental and clinical reports with consistent results in time, the evidence is supportive for level II (convincing evidence of beneficial action).^(53,54)

This is supported by the fact that clinical expertise and patient's choices have a much more central place in evidence-based medicine than some years ago.⁽⁵⁰⁾

The *American Heart Association Classification* asks for RCT's for the levels I and II. Prospective, controlled, but not randomized cohort studies are level III. In the light of the above mentioned data, we place hyperbaric oxygen as an adjunct to surgery and antibiotics in the treatment of gas gangrene in Class IIb (fair to good evidence provides support to Class IIa (very good evidence provides support)).

References

1. Stevens DL. The pathogenesis of clostridial myonecrosis. *Int J Med Microbiol* 2000, Oct; 290(4-5):497-502.
2. Lucey BP, Hutchins GM. William H. Welch and the discovery of *Bacillus Welchii*. *Arch Path Lab Med* 2004; 128 (10): 1193-95.
3. Weinstein L, Barza, MA. Medical intelligence. Current concepts: gas gangrene. *N Engl J Med* 1973; 289:1129B1131.
4. Bakker, DJ. Clostridial myonecrosis. In: Bakker DJ, Cramer FS eds. *Hyperbaric Surgery: Perioperative Care*. Falstaff, Az. Best Publ Company, 2002, 283-316.
5. Williamson ED, Titball RW. A genetically engineered vaccine against alpha-toxin of *Clostridium perfringens* protects mice against experimental gas gangrene. *Vaccine* 1993; 11(12): 1253B1258.
6. Stevens DL, Titball RW, Jepson M et al. Immunization with the C-Domain of α -Toxin prevents lethal infection, localizes tissue injury, and promotes Host response to challenge with *Clostridium Perfringens*. *J Inf Dis* 2004; 190: 767-73.
7. McLeod JW. Variations in the periods of exposure to air and oxygen necessary to kill anaerobic bacteria. *Acta Pathol Microbiol Scand* 1930; 3(suppl): 255.
8. Shimizu T, Ohtani K, Hirakawa H et al. Complete genome sequence of *Clostridium perfringens*, an anaerobic flesh-eater. *Proc Natl Acad Sci* 2002; 99 (2): 996-1001.
9. MacLennan JD. The histotoxic clostridial infections of man. *Bacteriol Rev* 1962; 26:177B276.
10. Hitchcock CR, Demello FJ, Haglin JJ. Gangrene infection: New approaches to an old disease. *Surg Clin N Am* 1975; 55: 1043B1410.
11. Heimbach RD. Gas gangrene. In: Kindwall EP, ed. *Hyperbaric medicine practice*. Flagstaff, AZ: Best Publishers, 1994: 373B394.
12. Titball RW, Naylor CE, Basak AK. The *Clostridium* α -Toxin. *Anaerobe* 1999; 5 (2): 51-64.
13. Willis AT. *Clostridia of wound infection*. London: Butterworth, 1969:490.
14. Stevens DL, Bryant AE, Adams K, Mader JT. Evaluation of therapy with hyperbaric oxygen for experimental infection with *Clostridium perfringens*. *Clin Infect Dis* 1993; 17: 231B237.
15. Stevens DL, Troyer BE, Merrick DT et al. Lethal effects and cardiovascular effects of purified alpha- and theta-toxins from *Clostridium perfringens*. *J Inf Dis* 1988; 157: 272- 279.
16. Stevens DL, Bryant AE. The Role of Clostridial Toxins in the Pathogenesis of Gas Gangrene. *Clin Inf Dis* 2002; 35:P S93-S100.

17. Awad MM, Bryant AE, Stevens DL, Rood JJ. Virulence studies on chromosomal alpha-toxin and theta-toxin mutants constructed by allelic exchange provide genetic evidence for the essential role of alpha-toxin in *C. perfringens*-mediated gas gangrene. *Mol Biol* 1995; 15(2): 191-202.
18. Eaton JT, Naylor CE, Howells AM et al. Crystal structure of the *C. Perfringens* alpha-toxin with the active site closed by a flexible loop region. *J Mol Biol* 2002; 319(2): 275-281.
19. Stevens DL, Tweten RK, Awad MM et al. Clostridial gas gangrene: Evidence that alpha- and theta-toxins differentially modulate the immune response and induce acute tissue necrosis. *J Inf Dis* 1997; 176: 189-195.
20. Van Unnik AJM. Inhibition of toxin production in *Clostridium perfringens* in vitro by hyperbaric oxygen. *Antonie Leeuwenhoek Microbiol* 1965; 31:181B186.
21. Kaye D. Effect of hyperbaric oxygen on Clostridia in vitro and in vivo. *Proc Soc Exp Biol Med* 1967; 124:360B366.
22. Hill GB, Osterhout S. Experimental effects of hyperbaric oxygen on selected clostridial species I in vitro studies and II in vivo studies in mice. *J Infect Dis* 1972; 125:17B35.
23. Muhvich KH, Anderson LH, Mehm WJ. Evaluation of antimicrobials combined with hyperbaric oxygen in a mouse model of clostridial myonecrosis. *J Trauma* 1994; 36(1): 7-10.
24. Demello FJ, Hashimoto T, Hitchcock CR, Haglin JJ. The effect of hyperbaric oxygen on the germination and toxin production of *Clostridium perfringens* spores. In: Wada J, Iwa JT, eds. Proceedings of the fourth international congress on hyperbaric medicine. Baltimore, MD: Williams & Wilkins, 1970:270.
25. Schoemaker G. Oxygen tension measurements under hyperbaric conditions. In: Boerema I, Brummelkamp WH, Meijne NG, eds. Clinical application of hyperbaric oxygen. Amsterdam: Elsevier, 1964:330B335.
26. Kivisaari J, Niinikoski J. Use of silastic tube and capillary sampling technique in the measurement of tissue PO₂ and PCO₂. *Am J Surg* 1973; 125:623B627.
27. Sheffield PJ. Tissue oxygen measurements. In: Davis JC, Hunt TK, eds. Problem wounds: the role of oxygen. New York: Elsevier, 1988:17B51.
28. Nora PF, Mousavipour M, Laufman H. Mechanisms of action of high pressure oxygen in *Clostridium perfringens* exotoxin toxicity. In: Brown IW, Cox BG, eds. Hyperbaric medicine, publ 1404. Washington DC: National Academy of Science/National Research Council, 1966:544-551.
29. Nora PF, Mousavipour M, Mittlepunkt A, Rosenburg M, Laufman H. Brain as target organ in *Clostridium perfringens* cytotoxin. *Arch Surg* 1966; 92:243B246.
30. Roggentin T, Kleineidam RG, Majewski DM, et al. An immunoassay for the rapid and specific detection of three sialidase-producing clostridia causing gas gangrene. *J Immunol Methods* 1993; 157:125B133.
31. Scheven M. Nachweis von Clostridium perfringens aus mischinfiziertem Patientenmaterial mittels modifiziertem reversen CAMP-Test. *Z gesammte Hyg* 1991; 37(2): 90-91.
32. Demello FJ, Haglin JJ, Hitchcock CR. Comparative study of experimental *Clostridium perfringens* infection in dogs treated with antibiotics, surgery and hyperbaric oxygen. *Surgery* 1973; 73:936B941.
33. Kelley HG, Pace WG. Treatment of anaerobic infections in mice with hyperpressure oxygen. *Surg Forum* 1963; 14: 46B47.
34. Klopper PJ. Hyperbaric oxygen treatment after ligation of the hepatic artery in rabbits. In: Boerema I, Brummelkamp WH, Meijne NG, eds. Clinical application of hyperbaric oxygen. Amsterdam: Elsevier, 1964:31B35.
35. Schott H. Die Gasbrand Infektion (Prinzipien der Behandlung, Ergebnisse) Hefte Unfallheilk 1979; 138:179B86.
36. Nier H, Kremer K. Der Gasbrand-weiterhin ein diagnostisches und therapeutisches Problem. *Zentralbl Chir* 1984; 109: 402B17.
37. Pailler JL, Labeau F. La gangrene gazeuse: Une affection militaire? *Acta Chir Belg* 1986; 86:63.
38. Ertmann M, Havemann D. Behandlung des Gasödems. Ergebnisse einer retro- und prospektiven Analyse des unfallchirurgischen Krankenguts aus 20 Jahren. *Unfallchir* 1992; 95: 471 - 476.
39. Brummelkamp WH, Hogendijk J, Boerema I. Treatment of anaerobic infections (clostridial myositis) by drenching the tissues with oxygen under high atmospheric pressure. *Surgery* 1961; 49:299B302.
40. Brummelkamp WH. Considerations on hyperbaric oxygen therapy at three atmospheres absolute for clostridial infections type welchii. *Ann NY Acad Sci* 1965; 117:688B699.
41. Bakker DJ. The use of hyperbaric oxygen in the treatment of certain infectious diseases, especially gas gangrene and acute dermal gangrene. Wageningen, Holland: Drukkerij Veenman BV, 1984.
42. Hart GB, Lamb RC, Strauss M. Gas gangrene: a collective review. *J Trauma* 1983; 23:991B1000.

43. Holland JA, Hill GB, Wolfe WA. Experimental and clinical experience with hyperbaric oxygen in the treatment of clostridial myonecrosis. *Surgery* 1975; 77:75B85.
44. Van Zijl JJW. Discussion of hyperbaric oxygen. In: Brown IW, Cox BG, eds. *Proceedings of the third international conference on hyperbaric medicine*. Washington DC: National Academy of Sciences/National Research Council, 1966:552-554.
45. Heimbach RD. Gas gangrene. Review and update. *HBO Rev* 1980; 1:41-46.
46. Hirn M. Hyperbaric oxygen in the treatment of gas gangrene and perineal necrotizing fasciitis. A clinical and experimental study. *Eur J Surg (suppl)* 1993; 570:9B36.
47. Peirce EC II. Gas gangrene: A critique of therapy. *Surg Rounds* 1984; 7:17B25.
48. Kindwall EP chairman, Hyperbaric Oxygen Therapy Committee Report: UMS Report Number 5-23-77; 1977: 4.
49. Evidence-Based Medicine Working Group (G.Guyatt, chair). *Evidence-Based Medicine. A New Approach to Teaching the Practice of Medicine*. *JAMA* 1992; 268(17): 2420-5.
50. HaynesRB, Deveraux PH, Guyatt GH. Clinical expertise in the era of evidence-based medicine and patient treatment. *Evid Based Med* 2002; 7: 36-38.
51. Tibbles PM, Edelsberg JS. Hyperbaric oxygen therapy: A review. *New Eng J Med* 1996; 334(25): 1642-8.
52. Mitton C, Hailey D. Health Technology Assessment and Policy Decisions on Hyperbaric Oxygen Treatment. *Int J Techn Ass in Health Care* 1999; 15(4): 661-670.
53. Oriani G. Acute indications for hyperbaric oxygen therapy-final report. In: Marroni A, Mathieu D, Wattel F (eds). *The ECHM Collection 2005*, vol. 1: 51-52. Best Publ Company, Flagstaff, Az. Report of the First Consensus Conference of the European Committee for Hyperbaric Oxygen, Lille 1994.
54. Wattel F (chair). Recommendations of the Jury of the 7th European Consensus Conference on Hyperbaric Medicine. Anaerobic and aerobic infections including gas gangrene. Type I Recommendation, level C. Lille University Hospital Publication 2004(available through the ECHIM secretariat).

